organic compounds

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3-(1-Pyridinio)propanesulfonate and 3-(benzyldimethylammonio)propanesulfonate monohydrate

Katarzyna D. Koclega,^{a,b} Maksymilian Chruszcz,^a Anna Gawlicka-Chruszcz,^c Marcin Cymborowski^a and Wladek Minor^a*

^aUniversity of Virginia, Department of Molecular Physiology and Biological Physics, 1340 Jefferson Park Avenue, Charlottesville, VA 22908, USA, ^bTechnical University of Lodz, Faculty of Biotechnology and Food Sciences, Institute of Technical Biochemistry, Stefanowskiego 4/10, 93-923 Lodz, Poland, and ^cHKL Research Inc., 310 Old Ivy Way, Charlottesville, VA 22903, USA Correspondence e-mail: wladek@iwonka.med.virginia.edu

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3-(1-Pyridinio)propanesulfonate, $C_8H_{11}NO_3S$, and 3-(benzyldimethylammonio)propanesulfonate monohydrate, $C_{12}H_{19}$ - $NO_3S \cdot H_2O$, used as additives during protein refolding and crystallization, both crystallize in the monoclinic system in the $P2_1/c$ space group, with one molecule (or one set of molecules) per asymmetric unit. The solvent water molecule present in the second crystal structure results in the formation of a dimer through hydrogen bonds. The conformation of the propanesulfonate moiety is similar in both structures.

Comment

Non-detergent sulfobetaines (NDSBs), to which the title compounds belong, are zwitterionic molecules. NDSB-201 or 3-(1-pyridinio)propanesulfonate, (I), and NDSB-256 or 3-(benzyldimethylammonio)propanesulfonate, of which the monohydrate structure, (II), is reported here, are members of a larger family of compounds that was mainly developed to facilitate protein solubilization, as well as for improvement of protein stability (Vuillard et al., 1995). It was discovered that compounds from this group are very useful during protein refolding (Goldberg et al. 1995; Vuillard et al., 1998; Expert-Bezançon et al. 2003; Swope Willis et al., 2006) and purification (Vuillard et al., 1995). NDSBs prevent protein aggregation and are used as additives to protein solutions in isoelectric focusing. Recently, Collins et al. (2006) demonstrated the usefulness of NDSB-201 in differential scanning calorimetry. The properties of NDSBs with respect to protein solutions have also been noticed by protein crystallographers (Vuillard et al., 1994, 1996). During crystallization, a protein has to be stable in highly concentrated solution for a prolonged period of time, and the presence of chemicals preventing the

formation of an amorphous precipitate could be crucial for the success of a crystallization experiment.



NDSB-201 and NDSB-256 crystallize in the $P2_1/c$ space group, with one molecule or one set of molecules per asymmetric unit (Figs. 1 and 2). Both compounds have aromatic rings that influence packing in the crystal structures. Weak interactions also play an important role in crystal packing, especially in the case of the structure of (I); the contacts between H and O atoms that are at least 0.3 Å shorter than the sum of the van der Waals radii are listed in Table 1. In the case of (II), hydrogen bonds mediated by water molecules are important for the packing (Table 2). The NDSB-256 molecules in the structure of (II) form dimers through hydrogen bonds involving water molecules (Fig. 3). The hydrogen-bonding pattern corresponds to an $R_4^4(12)$ motif, as described by Bernstein et al. (1995). Atom O1, which does not form hydogen bonds with water molecules, is involved in short contacts with benzyl atom H6B and atom H9 from the aromatic ring. The distances H6B···O1($-x, \frac{1}{2} + y, \frac{1}{2} - z$) and H9···O1(1 + x, y, z) are 2.35 and 2.48 Å, respectively, while the distances $C6 \cdots O1(-x, \frac{1}{2} + y, \frac{1}{2} - z)$ and $C9 \cdots O1(1 + x, y, \frac{1}{2} - z)$ z) are 3.262 (2) and 3.167 (2) Å, respectively. The angles C6– $H6B \cdots O1(-x, \frac{1}{2} + y, \frac{1}{2} - z)$ and $C9 - H9 \cdots O1(1 + x, y, z)$ are 156 and 131°, respectively.

NDSBs with a three-carbon bridge between the S and N atoms (sulfopropyl non-detergent betaines) have been found to be superior for work with proteins (Vuillard *et al.*, 1995). It was proposed that a sulfopropyl NDSB may adopt a cyclic conformation, with a six-atom ring and an ionic link between N⁺ and SO₃⁻ in solution. The resulting hydrocarbon cluster might take part in hydrophobic protein–protein interactions. Our results show that for NDSB-201 and NDSB-256, such a conformation of the sulfopropyl moiety is not observed in the crystal structures, but of course it cannot be concluded that at least some of the molecules do not adopt the cyclic conformation in solution. In the crystal structures reported by us, the torsion angles S1–C1–C2–C3 and C1–C2–C3–N1 are –178.3 (1) and 171.3 (1)°, respectively, for NDSB-201, while for NDSB-256 they are 178.8 (1) and 169.2 (1)°.



Figure 1

The molecular structure of compound (I). Displacement ellipsoids are drawn at the 50% probability level.

In the Cambridge Structural Database (CSD, Version 5.27, update of January 2006; Allen, 2002), there are 12 structures with the sulfopropyl moiety attached to an N atom, forming ternary or quaternary amines. In the structures reported in the CSD, there is also no example in which the cyclic conformation of the ⁺N-CH₂-CH₂-CH₂-SO₃⁻ fragment is observed. It is also quite surprising that, although NDSB molecules are quite often used during protein refolding, there are only a few structures in the Protein Data Bank (PDB; Berman et al., 2000) for which the use of non-detergent sulfobetaines is reported in REMARK 280 (REMARK 280 contains information about crystals, solvent content and crystallization conditions). A search of the PDB in September 2006 revealed that for only six structures were NDSBs used for crystallization. NDSB-195 was used in two cases (PDB codes 2AUW and 2G4B), NDSB-201 was used in three cases (PDB codes 1NAX, 1UA2 and 2FGC), and the usage of NDSB-256 was reported in only one case (PDB code 2F96). It was noted that the application of NDSB-201 (Lolli et al., 2004) helped to prevent excessive nucleation and promoted crystal growth. Most probably the influence of compounds from the NDSB family in protein solution is similar to that observed in the case of arginine (Baynes & Trout, 2004) and NDSBs may be treated as 'neutral crowder' additives (Baynes & Trout, 2004).



Figure 2

The molecular structure of compound (II). Displacement ellipsoids are drawn at the 50% probability level.



Figure 3

Hydrogen bonds (dashed lines) in the crystal structure of (II). Labelled and unlabelled molecules are related by the symmetry code (-x, 1 - y, -z).

Both NDSB-201 and NDSB-256 were purchased from Anatrace. Crystallization was performed at room temperature and the crystals used for X-ray diffraction experiments were obtained by slow evaporation; NDSB-201 was crystallized from a 1:1 mixture of methanol and 70% ethanol, while NDSB-256 was crystallized from 10% propionic acid.

Compound (I)

Crystal data $C_8H_{11}NO_3S$ $M_r = 201.24$ Monoclinic, $P2_1/c$ a = 5.699 (1) Å b = 7.428 (1) Å c = 20.053 (2) Å $\beta = 93.384$ (7)° V = 847.4 (2) Å³ Data collection Rigaku R-AXIS RAPID

diffractometer ω scans with χ offset Absorption correction: multi-scan (Otwinowski *et al.*, 2003) $T_{\min} = 0.27, T_{\max} = 0.38$

Refinement

Refinement on F^2
$R[F^2 > 2\sigma(F^2)] = 0.030$
$wR(F^2) = 0.082$
S = 1.05
1641 reflections
163 parameters
All H-atom parameters refined

Z = 4 D_x = 1.577 Mg m⁻³ Cu K α radiation μ = 3.20 mm⁻¹ T = 103 (2) K Prism, colourless 0.5 × 0.4 × 0.3 mm

78761 measured reflections 1641 independent reflections 1636 reflections with $I > 2\sigma(I)$ $R_{\text{int}} = 0.040$ $\theta_{\text{max}} = 72.2^{\circ}$

$w = 1/[\sigma^2(F_o^2) + (0.0455P)^2]$	
+ 0.5835P]	
where $P = (F_0^2 + 2F_c^2)/3$	
$(\Delta/\sigma)_{\rm max} < 0.001$	
$\Delta \rho_{\rm max} = 0.41 \text{ e } \text{\AA}^{-3}$	
$\Delta \rho_{\rm min} = -0.45 \ {\rm e} \ {\rm \AA}^{-3}$	
Extinction correction: SHELXL92	7
(Sheldrick, 1997)	
Extinction coefficient: 0.0253 (12)	

Table 1

Hydrogen-bond geometry (Å, °) for (I).

$D - H \cdot \cdot \cdot A$	D-H	$H \cdots A$	$D \cdots A$	$D - H \cdots A$
$\begin{array}{c} C4 - H4 \cdots O1^{i} \\ C3 - H3A \cdots O3^{i} \\ C7 - H7 \cdots O2^{ii} \\ C5 - H5 \cdots O1^{iii} \end{array}$	0.94 (2)	2.26 (2)	3.175 (2)	165.6 (16)
	0.996 (19)	2.397 (19)	3.347 (2)	159.2 (14)
	0.93 (2)	2.41 (2)	3.155 (2)	136.5 (16)
	0.96 (2)	2.41 (2)	3.206 (2)	139.8 (16)

Symmetry codes: (i) $-x, y + \frac{1}{2}, -z + \frac{1}{2}$; (ii) $x + 1, -y + \frac{3}{2}, z + \frac{1}{2}$; (iii) $x, -y + \frac{3}{2}, z + \frac{1}{2}$.

Compound (II)

Crystal data

 $C_{12}H_{19}NO_3S \cdot H_2O$ Z = 4

 $M_r = 275.37$ $D_x = 1$

 Monoclinic, P_{21}/c Cu Ka

 a = 12.628 (1) Å
 $\mu = 2.2$

 b = 11.209 (1) Å
 T = 10

 c = 9.982 (1) Å
 Block,

 $\beta = 107.260$ (4)°
 0.45 ×

 V = 1349.3 (2) Å³
 V

Data collection

Rigaku R-AXIS RAPID

diffractometer

 ω scans with χ offset

Absorption correction: multi-scan (Otwinowski *et al.*, 2003) $T_{\min} = 0.710, T_{\max} = 0.780$ Z = 4 $D_x = 1.356 \text{ Mg m}^{-3}$ Cu Ka radiation $\mu = 2.21 \text{ mm}^{-1}$ T = 103 (2) KBlock, colourless $0.45 \times 0.15 \times 0.11 \text{ mm}$

36458 measured reflections 2600 independent reflections 2449 reflections with $I > 2\sigma(I)$ $R_{\text{int}} = 0.057$ $\theta_{\text{max}} = 72.1^{\circ}$

organic compounds

Refinement

Refinement on F^2	$w = 1/[\sigma^2(F_0^2) + (0.0464P)^2]$
$R[F^2 > 2\sigma(F^2)] = 0.039$	+ 0.8011P]
$wR(F^2) = 0.102$	where $P = (F_0^2 + 2F_c^2)/3$
S = 1.08	$(\Delta/\sigma)_{\rm max} < 0.001$
2600 reflections	$\Delta \rho_{\rm max} = 0.36 \text{ e} \text{ Å}^{-3}$
172 parameters	$\Delta \rho_{\rm min} = -0.47 \ {\rm e} \ {\rm \AA}^{-3}$
H atoms treated by a mixture of	Extinction correction: SHELXL97
independent and constrained	(Sheldrick, 1997)
refinement	Extinction coefficient: 0.0169 (8)

Table 2

Hydrogen-bond geometry (Å, °) for (II).

$D - H \cdot \cdot \cdot A$	D-H	$H \cdot \cdot \cdot A$	$D \cdot \cdot \cdot A$	$D - H \cdots A$
$O4-H1\cdots O3^{i}$ $O4-H2\cdots O2$	0.83 (3) 0.87 (3)	2.00 (3) 2.05 (3)	2.812 (2) 2.921 (2)	167 (3) 178 (3)

Symmetry code: (i) -x, -y + 1, -z.

All H atoms in NDSB-201, (I), were located in a difference map and their positional and isotropic displacement parameters were refined. In the case of NDSB-256 monohydrate, (II), water H atoms were located in a difference map and their positional and isotropic displacement parameters were refined. All other H atoms were included in the refinement in calculated positions and refined using a riding-model approximation, with C–H = 0.93 (aromatic), 0.96 (CH₃) or 0.97 Å (CH₂), and with $U_{iso}(H) = 1.2U_{eq}(C)$ for aromatic CH and CH₂, or 1.5 $U_{eq}(C)$ for CH₃ hydrogens.

For both compounds, data collection: *HKL-2000* (Otwinowski & Minor, 1997); cell refinement: *HKL-2000*; data reduction: *HKL-2000*; program(s) used to solve structure: *HKL-3000SM* (Minor *et al.*, 2006) and *SHELXS97* (Sheldrick, 1990); program(s) used to refine structure: *HKL-3000SM* and *SHELXL97* (Sheldrick, 1997); molecular graphics: *HKL-3000SM*, *ORTEPIII* (Burnett & Johnson, 1996) and *ORTEP-3* (Farrugia, 1997); software used to prepare material for publication: *HKL-3000SM*.

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References

- Allen, F. H. (2002). Acta Cryst. B58, 380-388.
- Baynes, B. M. & Trout, B. L. (2004). Biophys. J. 87, 1631-1639.
- Berman, H. M., Westbrook, J., Feng, Z., Gilliland, G., Bhat, T. N., Weissig, H., Shindyalov, I. N. & Bourne, P. E. (2000). Nucleic Acids Res. 28, 235– 242.
- Bernstein, J., Davis, R. E., Shimoni, L. & Chang, N.-L. (1995). Angew. Chem. Int. Ed. Engl. 34, 1555–1573.
- Burnett, M. N. & Johnson, C. K. (1996). ORTEPIII. Report ORNL-6895. Oak Ridge National Laboratory, Tennessee, USA.
- Collins, T., D'Amico, S., Georlette, D., Marx, J. C., Huston, A. L. & Feller, G. (2006). Anal. Biochem. 352, 299–301.
- Expert-Bezançon, N., Rabilloud, T., Vuillard, L. & Goldberg, M. E. (2003). Biophys. Chem. 100, 469–479.
- Farrugia, L. J. (1997). J. Appl. Cryst. 30, 565.
- Goldberg, M. E., Expert-Bezançon, N., Vuillard, L. & Rabilloud, T. (1995). *Fold. Des.* **1**, 21–27.
- Lolli, G., Lowe, E. D., Brown, N. R. & Johnson, L. N. (2004). Structure, 12, 2067–2079.
- Minor, W., Cymborowski, M., Otwinowski, Z. & Chruszcz, M. (2006). Acta Cryst. D62, 859–866.
- Otwinowski, Z., Borek, D., Majewski, W. & Minor, W. (2003). Acta Cryst. A59, 228–234.
- Otwinowski, Z. & Minor, W. (1997). *Methods in Enzymology*, Vol. 276, *Macromolecular Crystallography*, Part A, edited by C. W. Carter Jr & R. M. Sweet, pp. 307–326. New York: Academic Press.
- Sheldrick, G. M. (1990). Acta Cryst. A46, 467-473.
- Sheldrick, G. M. (1997). SHELXL97. University of Göttingen, Germany.
- Swope Willis, M., Hogan, J. K., Prabhakar, P., Liu, X., Tsai, K., Wei, Y. & Fox, T. (2006). Protein Sci. 14, 1818–1826.
- Vuillard, L., Baalbaki, B., Lehmann, M., Nørager, S., Legrand, P. & Roth, M. (1996). J. Cryst. Growth, 168, 150–154.
- Vuillard, L., Braun-Breton, C. & Rabilloud, T. (1995). Biochem. J. 305, 337– 343.
- Vuillard, L., Rabilloud, T. & Goldberg, E. (1998). Eur. J. Biochem. 256, 128– 135.
- Vuillard, L., Rabilloud, T., Leberman, R., Berthet-Colominas, C. & Cusack, S. (1994). FEBS Lett. 353, 294–296.